Chemical Modification of the α-Mannosidase Inhibitor Mannostain A: Synthesis of a Potent Inhibitor 1L-(1,2,3,5/4)-5-Amino-4-O-methyl-1,2,3,4cyclopentanetetrol^[‡]

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Keywords: Cyclitols / Aminocyclopentitols / Glycosidase inhibitors / α -Mannosidase inhibitors / Mannostatin A analogues

Demethylthio-, S-demethyl-, and S-ethyl derivatives of the α mannosidase inhibitor, mannostasin A, were synthesized and evaluated for their inhibition of Jack bean α -mannosidase with the prime objective of elucidating the role of the methylthio group. All methylthio derivatives had significantly lowered inhibitory potentials. However, one mannostatin A analogue with a methoxyl instead of the methylthio group exhibited about twofold enhancement of the activity. The structure and inhibitory activity relationships of mannostatin A and related compounds are discussed in light of our results. (© Wiley-VCH Verlag GmbH & Co. KGaA, 69451 Weinheim, Germany, 2005)

Introduction

Identification of a potent and specific α-mannosidase inhibitor, mannostatin A^[2–10] [1, 1D-(1,2,3,4/5)-4-amino-5methylthio-1,2,3-cyclopentanetriol],[11] has stimulated us to develop new glycosidase inhibitors of the aminocyclopentitol type. These inhibitors are thought to act as transition state mimics of the glycopyranosyl cations that are postulated to be formed during hydrolysis of glycosides.[12,13] It appears rather difficult to correlate clearly the structures of known α-mannosidase inhibitors^[14] with the conformational features of the transition state of mannopyranosyl cations. Recently, Winkler and Holan^[15,16] proposed two conformational models (flap-up and -down) for mannopyranosyl cations, and suggested that comparison of the structure of mannostatin A to the former is pertinent, having a good overlap of the 1- and 2-hydroxy groups in 1 with the respective 3- and 2-hydroxy groups of the flap-up model (Figure 1).

Identification of the potent and specific inhibition of αmannosidase by mannostatin A (1) has led us to investigate novel glycosidase inhibitors of the 5-amino-1,2,3,4-cyclopentanetetrol type, with elucidation of their structure-inhibitory activity relationships. In a preceding paper, the synthesis and evaluation of the α -mannosidase inhibitory activity of three deoxy derivatives, 2-4, of 1 were described,[17,18] pointing to the clear significance of each hydroxy group (Figure 2). The 3-hydroxy groups of 1 and related compounds were found to be essential, conceivably

Part of this work was previously reported: Ref.[1]

Mannostatin A: 1 Mannopyranosyl cation

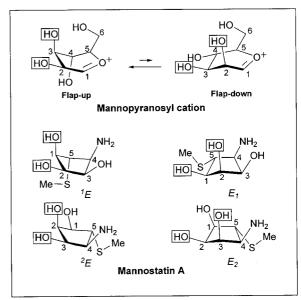


Figure 1. Mannostatin A (1) and mannopyranosyl cation. Comparison of preferred conformations (flap-up and -down) of the postulated transition-state mannopyranosyl cation with those of mannostatin A.

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Fax: +81-45-566-1789 E-mail: ogawa@bio.keio.ac.jp corresponding to the 2-hydroxy group of the mannopyranosyl cation. The amino groups located on the β-faces between the ring oxygen atoms and anomeric carbon atoms

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of these derivatives were also found to be essential. Thus, among the 24 known stereoisomers^[19] of 5-amino-1,2,3,4-cyclopentanetetrol, only **L-5** and **7**, and the corresponding 5-C-methyl derivatives,^[20] **DL-6** and **8**, were found to possess moderate inhibitory activity against Jack bean α -mannosidase (Table 1). Comparison of the activities of **L-5** and **L-9** suggests the role of the 3-hydroxy groups. All compounds containing cis-1,2-dihydroxy groups are likely to be matched with the 2- and 3-hydroxy groups of the mannopyranosyl cation. This consideration is in line with the findings that only mannostatin A isomer^[21] **D-10** and the 2,3-

SMe
$$H_2N$$
 OH H_2N OH

Figure 2. Three deoxy derivatives of mannostatin A and analogous aminocyclopentitol α -mannosidase inhibitors.

Table 1. Inhibitory activity (IC $_{50},\,\mu\text{M})$ against $\alpha\text{-mannosidase}$ (Jack beans); NI: no inhibition $<10^{-3}$ M.

Compound ^[a]	Inhibitory activity			
1	0.35			
2	28			
2 3	31			
4	NI			
D-5	110			
L-5	10			
L -6	83			
7	29			
8	56			
D -9	NI			
ь-9	250			
D-10	17 ^[b]			
11	11			
12	36			
13	5			
14	0.16			
15	NI			
16	NI			

[a] Only inhibitory activity against α -mannosidase is listed. [b] The K_i value was shown. [21]

diepimer^[22] are moderate α -mannosidase inhibitors, and no other isomers^[21,23] lacking vicinal *cis*-hydroxy groups show inhibitory activity.

In this paper we describe the details of chemical modification^[1] of the methylthio functionality of mannostatin A (1), with the focus on the synthesis and evaluation of the inhibitory activity of demethylthio (11), de-S-methyl (12), and ethylthio (13) derivatives (Figure 3). Furthermore, in order to assess the role of the sulfur atom of 1, attempts were made to replace the methylthio group with a methoxy group, preparing 4-O-methyl-, 1,4-di-O-methyl-, and epi-4-O-methylaminocyclopentanetetrols (14, 15 and 16), which led to finding a very strong α -mannosidase inhibitor, 14, comparable to the parent 1.

Figure 3. Chemical modification of the methylthio functionality of mannostatin A.

Results and Discussion

Reaction of the 2,3-*O*-cyclohexylidene derivative^[24,25] of (1,2,3,4,5/0)-5-acetamido-1,2,3,4-cyclopentanetetrol with (2*R*)-*O*-acetylmandelic acid in the presence of DCC and DMAP in CH₂Cl₂ diastereoselectively afforded 1*R*(2*R*)-*O*-acetylmandelate^[26] L-17 (56%) together with the (1*S*) ester D-17 (8%) (Figure 4). Compound L-17 was converted into the phenylthiocarbonyl ester 18 (71%) by treatment with DMAP (6 equiv.) and phenyl chlorothionocarbonate (5 equiv.) in CH₃CN at room temperature. Reaction of 18 with tributyltin hydride in the presence of AIBN gave the deoxy derivative 20 (71%). Deprotection of 20 with 2 M HCl at 80 °C, followed by conventional acetylation, gave the tetra-*N*,*O*-acetyl derivative 11a (ca. 100%), the structure of which was characterized on the basis of its ¹H NMR spectrum.

Compound L-17 was treated with triflic anhydride in pyridine/CH₂Cl₂ at -15 °C, and the resulting triflate 19 was then subjected to nucleophilic substitution with potassium thioacetate/18-crown-6 ether in dry benzene to give the acetylthio derivative 21 (61%). Acid hydrolysis of 21 followed by acetylation gave the penta-*N*,*O*,*S*-acetyl derivative 12a (ca. 100%). In addition, 21 was de-*S*-acetylated with methanolic sodium methoxide and the resulting crude thiol was treated with iodoethane to give the ethylthio derivative 22

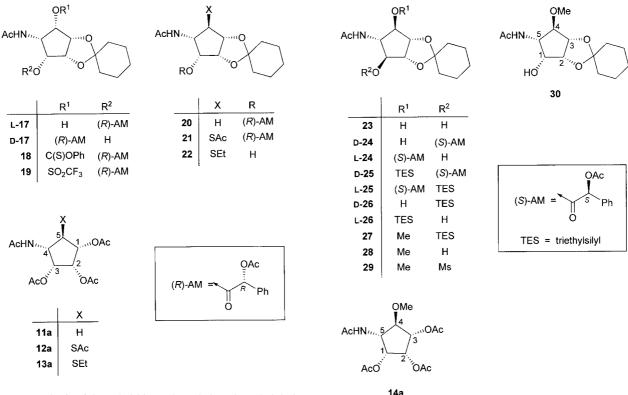


Figure 4. Synthesis of demethylthio, S-demethyl, and S-ethyl derivatives of mannostatin A.

(ca. 100%), which was purified by conversion into the tetra-

Figure 5. Synthesis of 1L-(1,2,3,5/4)-5-amino-4-*O*-methyl-1,2,3,4-cyclopentanetetrol (14).

N,O-acetyl derivative **13a** (ca. 100%). Diastereoselective acylation of the 2,3-O-cyclohexylidene derivative^[26] of (1,4/2,3,5)-5-acetamido-1,2,3,4-cyclopentanetetrol (23) with (2S)-O-acetylmandelic acid afforded a mixture (ca. 4:1) of the esters L-24 and D-24, which was directly treated with triethylsilyl triflate in CH₂Cl₂ to give a separable mixture of the triethylsilylates L-25 (77%) and D-25 (23%) (Figure 5). Compound L-25 was de-O-acylated under Zemplén conditions to give 26, which was subsequently treated with iodomethane/Ag₂O in acetonitrile to give the methyl ether 27 (ca. 100%). Desilylation of 27 with tetrabutylammonium fluoride in THF to give 28 (69%), was followed by conventional transformation into the mesylate 29. Crude mesylate 29 was hydrolyzed by heating in 80% aq. DMF at 110 °C to give, through neighboringgroup participation, the 4-epimeric alcohol 30 (76% overall yield). Acid hydrolysis of 30 and successive acetylation gave the tetra-N,O-acetyl derivative **14a** (53%).

The alcohol L-26 derived from D-25 was mesylated and the resulting crude sulfonate was treated with sodium acetate in aqueous DMF to give the 4(1)-epimer 31 (80%) of 23 (Figure 6). The structure of 31 was confirmed by the ¹H NMR spectrum of its di-*O*-acetyl derivative 32. Compound 31 was then methylated in a conventional manner to give the dimethyl ether 33 (68%), which was converted conventionally into the tri-*N*,*O*-acetyl derivative 15a (86%).

Compound D-17 was silylated and deacylated to give 34 (68%), and then methylated to give the methyl ether 35

Figure 6. Synthesis of the 1-O-methyl derivative 15a and 4-epimer 16a of 14a.

(97%). Conventional deprotection of **35** followed by acetylation gave the tetra-*N*,*O*-acetyl derivative **16a** (87%).

Mannostatin A analogues 11–13 were prepared by treatment of the corresponding peracetyl derivatives 11a–13a with 2 M hydrochloric acid at 80 °C, and purified on a col-

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umn of Dowex-50 W×2 (H⁺) resin with 1% aqueous ammonia as the eluent. The analogues **14–16** were also obtained by hydrolysis of **14a–16a** with 1 M aqueous Ba(OH)₂ at 90 °C, followed by a similar purification. The free bases thus obtained were directly subjected to assays of α -mannosidase inhibition.

Biological Assay

Listed in Table 1 are the Jack bean α-mannosidase inhibitory activities^[27] for newly prepared compounds 11-16, together with those of deoxy- (2-4), C-methyl- (6 and 8), and hydroxymethyl (9 and 10) aminocyclopentitol derivatives (Table 1). The moderate inhibitor 4-amino-1,2,3-cyclopentanetriol (11) appeared to have the minimum core structure for exhibiting inhibitory activity against α-mannosidase, suggesting that possible enhancement of activity could be achieved by chemical modification, for example, through Nsubstitution with alkyl or phenylalkyl functionalities.^[28] Incorporation of the appropriate functionalities at the 5-carbon atom of 11 with methylthio and methoxy groups, as shown in compounds 1 and 14, increased the activity dramatically. In contrast, derivatives having hydrophilic substituents at C-5, such as L-5 and 7, exhibited decreased activity, indicating that the presence of a hydrophobic region of space around C-5 is essential. Furthermore, the fact that the 4-epimer (16) of 14 completely lacked activity suggests that substituents located on the β -face at C-5 are important. It became apparent that the more the structures resembled mannostatin A (1), the greater their inhibitory potential. Farr^[29] reported the synthesis of a strong mannosidase inhibitor, amino(hydroxymethyl)cyclopentanetriol 36, with a 1L-(1,2,4/3,5N) configuration, and demonstrated good overlap between 36 and the mannopyranosyl cation by molecular modeling (Figure 1 and Table 2).^[16] Recently, Jäger^[30] described the synthesis of two stereoisomers, 37 and 38, having 1L-(1,2,4,5N/3) and 1L-(1,2/3,4,5N) configurations, respectively. From a structural viewpoint, the cis-vicinal hy-

Table 2. Present data for inhibitory activity (IC₅₀ [μM]) of mannostatin A (1) and 14, and those reported for isomeric amino(hydroxymethyl)cyclopentanetriols 36–38 against α-mannosidase (Jack beans) (reference compound 1: IC₅₀ = 0.02; 0.07 μM].

Compd.	\mathbb{R}^1	\mathbb{R}^2	\mathbb{R}^3	\mathbb{R}^4	X	Y	IС ₅₀ [µм]
1	ОН	Н	Н	SMe	NH_2	Н	0.32
14	OH	Η	Н	OMe	NH_2	Н	0.16
36	Η	OH	CH ₂ OH	Н	Н	NH_3Br	0.39
37	Η	OH	CH ₂ OH	Н	NH ₃ Br	Н	0.17
38	Н	ОН	H	CH ₂ OH	Н	NH ₃ Br	5.8

droxy groups adjacent to the amino group should match with the 2- and 3-hydroxy groups of the cation adopting the flap-up conformation, as roughly depicted in Figure 1. Although studies^[31] on a moderate α-mannosidase inhibitor, deoxymannonojirimycin, demonstrated that 5-hydroxymethyl branching is not indispensable for inhibitory action, we found the strong inhibitory activity of compounds 36 and 37 to depend on the presence of 4-hydroxymethyl groups, matching the 5-hydroxymethyl group of the mannopyranosyl cation. In the cases of compounds 1 and 14, the methylthio and methoxy groups seem to act as hydroxymethyl equivalents to provide a certain hydrophobic region of space, increasing their potency.

Conclusions

We obtained a strong α -mannosidase inhibitor amino-Omethylcyclopentanetriol 14, the activity of which was comparable to that of the parent 1, providing a lead compound for further development. The configuration of the hydrophobic S- or O-methyl functionality was shown to be very important, and the significance of a free 1-hydroxy group adjacent to the amino group was again verified by comparing the activities of the pairs 14 and 16, and 14 and 15, respectively. Strong inhibitory activity was demonstrated to be generated by an essential core structure composed of the 5-O-(or S-)methyl functionality and consecutive 1-, 2-, and 3-hydroxy, and 4-amino groups on the cyclopentane ring. Vicinal 2- and 3-hydroxy groups in a cis relationship match the 3- and 2-hydroxy groups of the postulated mannopyranosyl cation, and the amino functionality locates on the α- or β-face between pyranoid oxygen atom and anomeric carbon atoms. N-Substitution with normal or modified alkyl and phenylalkyl groups might improve inhibitory potential, as suggested by previous results.[28,30]

Experimental Section

General: Melting points: Mel-Temp capillary melting-point apparatus, uncorrected values. Specific rotations: Jasco DIP-370 polarimeter, 1-dm cells. IR spectra: Hitachi FT/IR-200 and BIORAD DIGITAL FT-65 spectrometers. ¹H NMR spectra: Jeol JNM EX-90 (90 MHz); Jeol GSX-270 f.t. (270 MHz), and Jeol Lambda-300 (300 MHz) spectrometers; solvent CDCl₃: internal tetramethylsilane (TMS) standard. CD₃OD: external acetone standard; D₂O: external acetone standard. Mass spectra: positive-ion electrospray ionization with a Micromass Zab Hybrid Spec Sector-TOF mass spectrometer. TLC: Silica Gel 60 GF (E. Merck Darmstadt); detection by charring with concentrated H₂SO₄. Column chromatography: Wakogel C-300 (silica gel, 300 mesh, Wako Chemical Osaka). Organic solutions, after drying with anhydrous Na₂SO₄, were concentrated at <50 °C under reduced pressure.

2,3-*O*-Cyclohexylidene Derivative of (1*R*,2*S*,3*R*,4*S*,5*S*)-5-Acetamido-1-*O*-[(2*R*)-2-*O*-acetylmandelyl]-4-*O*-phenoxythiocarbonyl-1,2,3,4-cyclopentanetetrol (18): To a solution of L-17^[26] (1.83 g, 4.1 mmol) in acetonitrile (18 mL) were added DMAP (3.0 g, 24.5 mmol, 6 equiv.) and phenyl chlorothioformate (2.83 mL, 20.5 mmol, 5 equiv.), and the reaction mixture was stirred at room temperature

for 3 h. The mixture was diluted with ethyl acetate (200 mL), the solution was washed with water, dried, and the solvents were evaporated. The residual product was purified by chromatography on a silica gel column (240 g, acetone/toluene, 1:10) to give **18** (1.69 g, 71%) as a syrup. TLC (acetone/toluene, 1:3): $R_{\rm f}=0.43$. [a] $_{\rm D}^{18}=-30$ (c=0.64, CHCl $_{\rm 3}$). $^{1}{\rm H}$ NMR (270 MHz, CDCl $_{\rm 3}$): $\delta=1.22-1.77$ (m, 10 H, C $_{\rm 6}$ H $_{\rm 10}$), 1.93, 2.21 (2 s, each 3 H, 2×Ac), 4.76 (dd, $J_{\rm 1,2}=5.1$, $J_{\rm 1,5}=5.8$ Hz, 1 H, 1-H), 4.82 (dd, $J_{\rm 3,4}=5.1$, $J_{\rm 2,3}=5.5$ Hz, 1 H, 3-H), 4.94 (ddd, $J_{\rm 4,5}=5.6$, $J_{\rm 1,5}=5.8$, $J_{\rm 5,NH}=9.2$ Hz, 1 H, 5-H), 5.01 (dd, $J_{\rm 1,2}=5.1$, $J_{\rm 2,3}=5.5$ Hz, 1 H, 2-H), 5.47 (dd, $J_{\rm 3,4}=5.1$, $J_{\rm 4,5}=5.6$ Hz, 1 H, 4-H), 6.01 [s, 1 H, PhCH(OAc)CO], 6.33 (d, $J_{\rm 5,NH}=9.2$ Hz, 1 H, NH), 7.12–7.59 (m, 10 H, 2×Ph) ppm. HRMS: calcd. for C $_{\rm 30}$ H $_{\rm 33}$ NO $_{\rm 9}$ S 583.1876; found 583.1871 [M $^{+}$].

1,2-O-Cyclohexylidene Derivative of (1S,2S,3R,4R)-4-Acetamido-3-O-[(2R)-2-O-acetylmandelyl]-1,2,3-cyclopentanetetrol (20): To a solution of AIBN (3.4 mg, 0.021 mmol, 0.3 equiv.) in toluene (0.5 mL) was added tributyltin hydride (55.4 μL, 0.21 mmol, 3 equiv.) dropwise, followed by a mixture of 18 (40 mg, 0.07 mmol) and toluene (3.5 mL). The reaction mixture was heated at reflux for 2 h, then concentrated to dryness, and the product was purified by chromatography on a silica gel column (10 g, acetone/toluene, 1:4.5) to give 20 (21 mg, 71%) as a syrup. TLC (acetone/toluene, 1:3): $R_f = 0.18$. $[a]_D^{22} = +21$ (c = 0.54, CHCl₃). ¹H NMR (300 MHz, CDCl₃): $\delta = 1.25-1.52$ (m, 10 H, C₆H₁₀), 1.98-2.04 (m, 2 H, 5a-H, 5b-H), 1.87, 2.20 (2 s, each 3 H, 2×Ac), 4.57 (m, 1 H, 4-H), 4.63 (ddd, $J_{1,5a}$ = 2.4, $J_{1,5b}$ = 5.4, $J_{1,2}$ = 6.1 Hz, 1 H, 1-H), 4.70 (dd, $J_{2,3}$ = 4.9, $J_{1,2}$ = 6.1 Hz, 1 H, 2-H), 4.87 (dd, $J_{2,3}$ = $J_{3,4}$ = 4.9 Hz, 1 H, 3-H), 5.99 [s, 1 H, PhCH(OAc)CO], 6.26 (d, $J_{4,NH}$ = 9.0 Hz, 1 H, NH), 7.40-7.52 (m, 5 H, Ph) ppm. HRMS: calcd. for C₂₃H₃₀NO₉ 431.1944; found 431.1949 [M+].

Tetra-*N*, *O*-acetyl-(1*S*,2*S*,3*R*,4*R*)-4-amino-1,2,3-cyclopentanetriol (11a): A mixture of 20 (8.2 mg, 19 μmol) and 2 M hydrochloric acid (1.0 mL) was heated at 80 °C for 1 h, and then concentrated to dryness. The residue was treated with acetic anhydride (0.5 mL) and pyridine (1 mL) at room temperature overnight. The reaction mixture was coconcentrated with toluene, and the residue was purified by chromatography on a column of silica gel (0.7 g, acetone/toluene, 1:2) to give 11a (5.7 mg, ca. 100%) as a syrup. TLC (acetone/toluene, 1:1): $R_{\rm f} = 0.39$. [a] $_{\rm f}^{\rm D5} = +16$ (c = 0.42, CHCl₃). $^{\rm 1}$ H NMR (300 MHz, CDCl₃): δ = 1.77 (m, 1 H, 5a-H), 2.01, 2.05, 2.07, 2.13 (4 s, each 3 H, 4×Ac), 2.66 (m, 1 H, 5b-H), 4.57 (m, 1 H, 4-H), 5.20–5.28 (m, 3 H, 1-H, 2-H, 3-H), 5.74 (d, $J_{4,\rm NH} = 8.5$ Hz, 1 H, NH) ppm. HRMS: calcd. for C₁₃H₂₀NO₇ 302.1239; found 302.1229 [M⁺].

Penta-*N*, *O*, *S*-acetyl-(1*R*, 2*R*, 3*R*, 4*S*, 5*S*)-4-amino-5-thio-1, 2, 3-cyclopentanetriol (12a): A mixture of $21^{[26]}$ (12.0 mg, 24 μmol) and 2 M hydrochloric acid (1.0 mL) was stirred at 80 °C for 1 h, and then coconcentrated with EtOH. The residue was acetylated in the usual manner and the product was purified by chromatography on a column of silica gel (0.7 g, acetone/toluene, 1:4) to give 12a (8.9 mg, ca. 100%) as a syrup. TLC (acetone/toluene, 1:1): $R_f = 0.48$. [a] $_D^{25} = +26$ (c = 0.37, CHCl $_3$). $_1^{1}$ H NMR (300 MHz, CDCl $_3$): $_3^{1}$ = 1.99, 2.03, 2.07, 2.15, 2.37 (5 s, each 3 H, 5×Ac), 3.91 (dd, $J_{1,5} = 6.8$, $J_{4,5} = 10.5$ Hz, 1 H, 5-H), 4.52 (ddd, $J_{3,4} = 4.4$, $J_{4,NH} = 8.5$, $J_{4,5} = 10.5$ Hz, 1 H, 4-H), 5.26 (dd, $J_{1,5} = 6.8$, $J_{1,2} = 7.1$ Hz, 1 H, 1-H), 5.40 (dd, $J_{2,3} = 3.9$, $J_{1,2} = 7.1$ Hz, 1 H, 2-H), 5.42 (dd, $J_{2,3} = 3.9$, $J_{3,4} = 4.4$ Hz, 1 H, 3-H), 5.79 (d, $J_{4,NH} = 8.5$ Hz, 1 H, NH) ppm. HRMS: calcd. for C₁₅H₂₄NO₇S 376.1066; found 376.1068 [M⁺].

1,2-O-Cyclohexylidene Derivative of (1R,2S,3R,4S,5R)-4-Acetamido-5-ethylthio-1,2,3-cyclopentanetriol (22): A solution of 21 (5.7 mg, 11 μ mol) in MeOH (0.2 mL) was treated with 1 μ methanolic sodium methoxide (17 μ L) at room temperature for 10 min,

and then iodoethane (9 µL, 1.1 mmol) was added. After stirring at room temperature for 1 h, the reaction mixture was concentrated and the residue was purified by chromatography on a column of silica gel (0.5 g, acetone/toluene, 1:3) to give **22** (3.6 mg, ca. 100%) as a syrup. TLC (acetone/toluene, 1:2): $R_{\rm f} = 0.27$. [a] $_{\rm D}^{22} = +1.7$ (c = 0.35, CHCl₃). 1 H NMR (300 MHz, CDCl₃): $\delta = 1.29$ (t, J = 7.3 Hz, 3 H, CH₂CH₃), 1.38–1.77 (m, 10 H, C₆H₁₀), 2.03 (s, 3 H, Ac), 2.70 (q, J = 7.3 Hz, 2 H, CH₂CH₃), 3.17 (dd, $J_{3,4} = 3.9$, $J_{4,5} = 8.8$ Hz, 1 H, 4-H), 4.16 (dd, $J_{1,5} = 4.4$, $J_{1,2} = 4.9$ Hz, 1 H, 1-H), 4.34 (ddd, $J_{1,5} = 4.4$, $J_{4,5} = 8.8$, $J_{5,\rm NH} = 9.0$ Hz, 1 H, 5-H), 4.51 (dd, $J_{3,4} = 3.9$, $J_{2,3} = 7.1$ Hz, 1 H, 3-H), 4.58 (dd, $J_{1,2} = 4.9$, $J_{2,3} = 7.1$ Hz, 1 H, 2-H), 6.08 (d, $J_{5,\rm NH} = 9.0$ Hz, 1 H, NH) ppm. HRMS: calcd. for C_{15} H₂₅NO₄S 315.1504; found 315.1506 [M $^+$].

Tetra-*N*,*O*-acetyl-(1*R*,2*R*,3*R*,4*S*,5*S*)-4-amino-5-ethylthio-1,2,3-cyclopentanetriol (13a): Compound 22 (6.0 mg, 19 μmol) was hydrolyzed as in the preparation of 11, and the product was acetylated in the usual manner. The product was purified by a column of silica gel (0.7 g, acetone/toluene, 1:2) to give 13a (6.9 mg, ca. 100%) as a syrup. TLC (acetone/toluene, 1:1): $R_{\rm f} = 0.40$. [a] $_{\rm D}^{25} = +15$ (c = 0.07, CHCl₃). 1 H NMR (300 MHz, CDCl₃): $\delta = 1.25$ (t, J = 7.3 Hz, 3 H, CH₂CH₃), 2.04, 2.06, 2.08, 2.11 (4 s, each 3 H, 4×Ac), 2.45 (br. q, J = 7.3 Hz, 2 H, CH₂CH₃), 3.17 (dd, $J_{1,5} = 5.6$, $J_{4,5} = 7.6$ Hz, 1 H, 5-H), 4.53 (ddd, $J_{3,4} = 5.6$, $J_{4,5} = 7.6$, $J_{4,\rm NH} = 8.8$ Hz, 1 H, 4-H), 5.16 (t, $J_{1,2} = J_{1,5} = 5.6$ Hz, 1 H, 1-H), 5.35 (dd, $J_{2,3} = 4.4$, $J_{3,4} = 5.6$ Hz, 1 H, 3-H), 5.42 (dd, $J_{2,3} = 4.4$, $J_{1,2} = 5.6$ Hz, 1 H, 2-H), 5.70 (d, $J_{4,\rm NH} = 8.8$ Hz, 1 H, NH) ppm. HRMS: calcd. for C₁₅H₂₄NO₇S 362.1273; found 362.1270 [M⁺].

2,3-O-Cyclohexylidene Derivatives of (1S,2S,3R,4R,5R)- and (1R,2R,3S,4S,5S)-5-Acetamido-1-O-[(2S)-2-O-acetylmandelyl]-4-O-triethylsilyl-1,2,3,4-cyclopentanetetrol (D-25 and L-25): A mixture of 1D- and 1L-(1,4/2,3,5)-5-acetamido-1-O-[(2S)-2-O-acetylmandelyl]-1,2,3,4-cyclopentanetetrol (D-24 and L-24, 336 mg, 0.752 mmol) derived (2S) from the diol 23, triethylsilyl triflate $(255 \, \mu L, 1.13 \, mmol, 1.5 \, equiv.)$, and 2,6-lutidine $(262 \, \mu L, 2.25 \, mmol, 3 \, equiv.)$ in dichloromethane $(3.5 \, mL)$ was stirred under argon at 0 °C for 90 min. To the reaction mixture was then added saturated aqueous NaHCO₃ $(60 \, mL)$, and the aqueous layer was thoroughly extracted with chloroform $(3 \times 20 \, mL)$. The extracts were dried, and the solvents were evaporated. The residue was purified by chromatography on a silica gel column $(40 \, g, \, ethyl \, acetate/toluene, \, 1:7)$ to give L-25 $(326 \, mg, \, 77\%)$ and D-25 $(94 \, mg, \, 23 \, \%)$, each as a syrup.

L-25: TLC (acetone/toluene, 1:4): $R_{\rm f} = 0.48$. $[a]_{\rm D}^{25} = +37$ (c = 1.14, CHCl₃). $^1{\rm H}$ NMR (300 MHz, CDCl₃): $\delta = 0.59$ [dd, J = 7.8, $J_{\rm gem} = 16.1$ Hz, 6 H, Si(C H_2 CH₃)₃], 0.93 [t, J = 7.8 Hz, 9 H, Si(CH₂C H_3)₃], 1.39–1.70 (m, 10 H, C₆H₁₀), 1.85, 2.18 (2 s, each 3 H, 2×Ac), 4.02 (dd, $J_{1,2} = 2.2$, $J_{1,5} = 4.4$ Hz, 1 H, 1-H), 4.12 (ddd, $J_{1,5} = 4.4$, $J_{4,5} = 4.6$, $J_{5,\rm NH} = 9.0$ Hz, 1 H, 5-H), 4.41 (dd, $J_{1,2} = 2.2$, $J_{2,3} = 6.6$ Hz, 1 H, 2-H), 4.57 (dd, $J_{3,4} = 2.2$, $J_{2,3} = 6.6$ Hz, 1 H, 3-H), 5.04 (dd, $J_{3,4} = 2.2$, $J_{4,5} = 4.6$ Hz, 1 H, 4-H), 5.63 (br. d, $J_{5,\rm NH} = 9.0$ Hz, 1 H, NH), 5.87 [s, 1 H, PhCH(OAc)CO], 7.36–7.46 (m, 5 H, Ph) ppm. HRMS: calcd. for C₂₇H₃₈NO₈Si 532.2367; found 532.2367 [M⁺].

D-25: TLC (acetone/toluene, 1:4): $R_{\rm f} = 0.44$. $[a]_{\rm D}^{25} = +28$ (c = 1.12, CHCl₃). ¹H NMR (300 MHz, CDCl₃): $\delta = 0.63$ [dd, J = 8.1, $J_{\rm gem} = 15.4$ Hz, 6 H, Si(C H_2 CH₃)₃], 0.96 [t, J = 8.1 Hz, 9 H, Si(CH₂C H_3)₃], 1.37–1.71 (m, 10 H, C₆H₁₀), 1.92, 2.17 (2 s, each 3 H, 2×Ac), 4.07 (m, 1 H, 1-H), 4.26 (m, 1 H, 5-H), 4.33 (m, 2 H, 2-H, 3-H), 5.10 (m, 1 H, 4-H), 5.72 (br. d, $J_{5,\rm NH} = 9.0$ Hz, 1 H, NH), 5.90 [s, 1 H, PhCH(OAc)CO], 7.37–7.48 (m, 5 H, Ph) ppm. HRMS: calcd. for C₂₇H₃₈NO₈Si 532.2367; found 532.2388 [M⁺].

2,3-O-Cyclohexylidene Derivative of (1R,2S,3S,4S,5R)-5-Acetamido-1-O-triethylsilyl-1,2,3,4-cyclopentanetetrol (D-26): A solution

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of the *S*-acetylmandelate **L-25** (159 mg, 0.28 mmol) in CH₂Cl₂ (1.6 mL) was treated with 1 M methanolic sodium methoxide (57 μL, 57 μM) at 0 °C for 3 h. The product was purified by chromatography on a column of silica gel (10 g, acetone/toluene, 1:2) to give the alcohol **D-26** (109 mg, ca. 100%) as a syrup. TLC (acetone/toluene, 1:2): $R_f = 0.41$. [a] $_0^{25} = +10$ (c = 1.65, CHCl $_3$). 1 H NMR (300 MHz, CDCl $_3$): $\delta = 0.69$ [dd, J = 8.1, $J_{\rm gem} = 15.9$ Hz, 6 H, Si(CH $_2$ CH $_3$) $_3$], 0.99 [t, J = 8.1 Hz, 9 H, Si(CH $_2$ CH $_3$) $_3$], 1.36–1.77 (m, 10 H, C $_6$ H $_1$ 0), 1.97 (s, 3 H, Ac), 3.79 (d, $J_{4,\rm OH} = 8.1$ Hz, 1 H, OH), 4.00–4.07 (m, 3 H, 1-H, 4-H, 5-H), 4.50 (m, 1 H, 2-H), 5.64 (m, 1 H, 3-H), 5.90 (m, 1 H, NH) ppm. HRMS: calcd. C $_{19}$ H $_{35}$ NO $_5$ Si 385.2285; found 385.2303 [M $^+$].

- **2,3-***O*-Cyclohexylidene Derivative of (1*S*,2*R*,3*R*,4*R*,5*S*)-5-Acetamido-1-*O*-triethylsilyl-1,2,3,4-cyclopentanetetrol (L-26): D-25 (149 mg, 0.26 mmol) was deacylated as in the preparation of D-26 to give L-26 (167 mg, ca. 100%) as a syrup. TLC (acetone/toluene, 1:2): $R_f = 0.41$. [a] $_5^{25} = -7.4$ (c = 1.0, CHCl₃). HRMS: calcd. for C₁₉H₃₆NO₅Si 386.2363; found 386.2354 [M⁺]. The ¹H NMR spectrum was found to be superimposable on that of D-26.
- 2,3-O-Cyclohexylidene Derivative of (1S,2S,3S,4S,5R)-5-Acetamido-1-O-methyl-4-O-triethylsilyl-1,2,3,4-cyclopentanetetrol (27): To a solution of D-26 (12.3 mg, 2 mmol) in acetonitrile (0.5 mL) were added silver oxide (11.4 mg, 48 µM) and iodomethane (0.2 mL, 3.19 mmol), and the mixture was refluxed for 6 h. An insoluble material was removed by filtration, and the filtrate was concentrated to dryness. The residue was purified by chromatography on a silica gel column (0.7 g, acetone/toluene, 1:7) to give the methyl ether 27 (12.7 mg, ca. 100%) as a syrup. TLC (acetone/toluene, 1:1): $R_f = 0.71$. $[a]_D^{25} = -0.1$ (c = 0.7, CHCl₃). ¹H NMR (300 MHz, CDCl₃): $\delta = 0.63$ [dd, J = 8.1, $J_{gem} = 14.9$ Hz, 6 H, Si(C H_2 CH₃)₃], 0.96 [t, J = 8.1 Hz, 9 H, Si(C H_2 CH₃)₃], 1.32–1.70 (m, 10 H, C₆H₁₀), 1.97 (s, 3 H, Ac), 3.42 (s, 3 H, Me), 3.71 (m, 1 H, 4-H), 4.06-4.11 (m, 2 H, 1-H, 5-H), 4.41 (m, 1 H, 2-H), 4.52 (dd, $J_{2,3} = 2.2$, $J_{3,4} =$ 6.6 Hz, 1 H, 3-H), 5.73 (br. d, $J_{5,NH}$ = 7.6 Hz, 1 H, NH) ppm. HRMS: calcd. for $C_{20}H_{37}NO_5Si$ 399.2441; found 399.2441 [M⁺].
- 2,3-O-Cyclohexylidene Derivative of (1R,2S,3R,4S,5S)-5-Acetamido-1-O-methyl-1,2,3,4-cyclopentanetetrol (28): A solution of 27 (83 mg, 0.21 mmol) in THF (1.6 mL) was treated with TBAF (59 µL, 0.25 mmol) at room temperature for 1.5 h, and then the reaction mixture was concentrated to dryness. The residue was dissolved in ethyl acetate (15 mL), and the solution was washed with 1 м hydrochloric acid, saturated aqueous NaHCO₃, and water, dried, and the solvents were evaporated. The residual product was purified by chromatography on a silica gel column (5 g, acetone/ toluene, 1:1) to give 28 (40.5 mg, 69%) as a syrup. TLC (acetone/ toluene, 1:1): $R_f = 0.34$. $[a]_D^{27} = +12$ (c = 0.82, CHCl₃). ¹H NMR (300 MHz, CDCl₃): $\delta = 1.31-1.75$ (m, 10 H, C₆H₁₀), 2.01 (s, 3 H, Ac), 3.48 (s, 3 H, Me), 3,62 (m, 1 H, 1-H), 4.03 (m, 2 H, 4-H, 5-H), 4.29 (m, 1 H, OH), 4.56 (br. s, 2 H, 2-H, 3-H), 5.99 (br. s, 1 H, NH) ppm. HRMS: calcd. for C₁₄H₂₃NO₅ 285.1576; found 285.1589 $[M^+].$
- **2,3-***O*-Cyclohexylidene Derivative of (1R,2R,3S,4R,5S)-5-*O*-Acetamido-4-*O*-methyl-1,2,3,4-cyclopentanetetrol (30): To a solution of **28** (16.5 mg, 58 µmol) in pyridine (0.4 mL) was added mesyl chloride (22.4 µL, 0.29 mmol) at 0 °C. The mixture was stirred at room temperature for 1 h, and then coconcentrated with toluene. The residue was diluted with CHCl₃ (15 mL), and the solution was washed thoroughly with water, dried, and the solvents were evaporated. The resulting crude mesylate **29** was dissolved in aqueous DMF (80%, 3.0 mL) and, after addition of sodium acetate (18 mg, 2.2 mmol), the mixture was stirred at 110 °C overnight. The reaction mixture was then coconcentrated with ethanol and toluene,

and the residue was purified by chromatography on a silica gel column (1.5 g, acetone/toluene, 1:3) to give the alcohol **30** (12.6 mg, 76%) as a syrup. TLC (acetone/toluene, 1:1): $R_{\rm f}=0.42$. $[a]_{\rm D}^{25}=-29~(c=0.8,{\rm CHCl_3})$. ¹H NMR (300 MHz, CDCl₃): $\delta=1.38-1.76$ (m, 10 H, C₆H₁₀), 2.03 (s, 3 H, Ac), 3.23 (d, $J_{\rm 4,OH}=3.7$ Hz, 1 H, OH), 3.43 (s, 3 H, Me), 3.68 (dd, $J_{\rm 1,2}=2.7$, $J_{\rm 1,5}=6.6$ Hz, 3 H, 1-H), 4.11 (ddd, $J_{\rm 4,OH}=3.7$, $J_{\rm 4,5}=4.6$, $J_{\rm 3,4}=4.9$ Hz, 1 H, 4-H), 4.41 (ddd, $J_{\rm 4,5}=4.6$, $J_{\rm 1,5}=6.6$, $J_{\rm 5,NH}=8.5$ Hz, 1 H, 5-H), 4.45 (dd, $J_{\rm 1,2}=2.7$, $J_{\rm 2,3}=7.1$ Hz, 1 H, 2-H), 4.58 (dd, $J_{\rm 3,4}=4.9$, $J_{\rm 2,3}=7.1$ Hz, 1 H, 3-H), 6.12 (br. d, $J_{\rm 5,NH}=8.5$ Hz, 1 H, NH) ppm. HRMS: m/z=285.1575 [M⁺]. C₁₄H₂₃NO₅ (285.1576).

- (1*R*,2*R*,3*S*,4*R*,5*R*)-5-Acetamido-1,2,3-*O*-acetyl-4-*O*-methyl-1,2,3,4-cyclopentanetetrol (14a): A mixture of 30 (31.4 mg, 0.11 mmol) and aqueous acetic acid (60%, 1.0 mL) was stirred at 60 °C for 5.5 h and then concentrated to dryness. The residual product was acetylated in the usual manner, and the product was purified by chromatography on a silica gel column (3 g, acetone/toluene, 1:3) to give the acetyl derivative 14a (19.2 mg, 53%) as a syrup. TLC (acetone/toluene, 1:1): $R_f = 0.32$. [a] $_D^{23} = -3.1$ (c = 0.5, CHCl₃). $_D^{14}$ NMR (300 MHz, CDCl₃): $_D^{14} = 2.04$, 2.08, 2.10 (3 s, 3 H, 6 H, 3 H, 4×OAc), 3.42 (s, 3 H, Me), 3.77 (t, $_{J_{3,4}} = _{J_{4,5}} = 4.6$ Hz, 1 H, 4-H), 4.57 (ddd, $_{J_{4,5}} = 4.6$, $_{J_{1,5}} = 6.1$, $_{J_{5,NH}} = 9.0$ Hz, 1 H, 5-H), 5.14 (t, $_{J_{2,3}} = _{J_{3,4}} = 4.6$ Hz, 1 H, 3-H), 5.34 (dd, $_{J_{1,2}} = 3.9$, $_{J_{1,5}} = 6.1$ Hz, 1 H, 1-H), 5.46 (dd, $_{J_{1,2}} = 3.9$, $_{J_{2,3}} = 4.6$ Hz, 1 H, 2-H), 5.82 (br. d, $_{J_{5,NH}} = 9.0$ Hz, 1 H, NH) ppm. HRMS: calcd. for $_{L_4}$ C₁₄ H₂₂NO₈ 332.1345; found 332.1371 [M⁺].
- 2,3-O-Cyclohexylidene Derivative of (1S,2R,3S,4R)-5-Acetamido-**1,2,3,4-cyclopentanetetrol (31):** Alcohol L**-26** (167 mg, 0.433 mmol) was mesylated as in the preparation of 29. During the above processing, when the reaction mixture was concentrated and coconcentrated with toluene, a partial removal of the triethylsilyl group was observed. The resulting crude mesylate was treated with sodium acetate (178 mg, 2.16 mmol) in aqueous 80% DMF (2.0 mL) at 110 °C overnight, and then the mixture was concentrated and coconcentrated with ethanol and toluene. The residue was purified by chromatography on a column of silica gel (10 g, acetone/toluene, 1:1) to give the diol 31 (94 mg, 80%) as a syrup. TLC (acetone/ toluene, 2:1): $R_f = 0.28$. $[a]_D^{21} = -28$ (c = 1.0, CHCl₃). ¹H NMR (300 MHz, CDCl₃): $\delta = 1.34-1.76$ (m, 10 H, C₆H₁₀), 2.03 (s, 3 H, Ac), 4.13–4.21 (m, 3 H, 3-H, 4-H, 5-H), 4.49 (d, $J_{1,2} = 7.1$ Hz, 1 H, 1-H), 4.65 (dd, $J_{2,3}$ = 4.9, $J_{1,2}$ = 7.1 Hz, 1 H, 2-H), 6.52 (br. s, 1 H, NH) ppm. HRMS: calcd. for C₁₃H₂₁NO₅ 271.1420; found 271.1413 [M⁺].
- **2,3-***O*-Cyclohexylidene Derivative of (1*S*,2*S*,3*R*,4*R*)-5-Acetamido-1,4-di-*O*-acetyl-1,2,3,4-cyclopentanetetrol (32): Diol 31 (91 mg, 0.34 mmol) was acetylated with acetic anhydride in pyridine in the usual manner, and the product was purified by chromatography on a column of silica gel (10 g, acetone/toluene, 1:4) to give the di-*O*-acetyl derivative 32 (119 mg, ca. 100%) as a syrup. TLC (acetone/toluene, 1:1): $R_{\rm f} = 0.50$. [a] $_{\rm D}^{23} = +38$ (c = 1.5, CHCl $_{\rm 3}$). 1 H NMR (300 MHz, CDCl $_{\rm 3}$): $\delta = 1.25$ –1.82 (m, 10 H, C $_{\rm 6}$ H $_{\rm 10}$), 1.99, 2.08, 2.16 (3 s, each 3 H, 3×Ac), 4.48 (dd, $J_{3,4} = 1.7$, $J_{2,3} = 6.3$ Hz, 1 H, 3-H), 4.54 (ddd, $J_{4,5} = 4.6$, $J_{1,5} = 5.1$, $J_{5,\rm NH} = 8.5$ Hz, 1 H, 5-H), 4.77 (dd, $J_{1,2} = 5.1$, $J_{2,3} = 6.3$ Hz, 1 H, 2-H), 5.11 (dd, $J_{3,4} = 1.7$, $J_{4,5} = 4.6$ Hz, 1 H, 4-H), 5.18 (t, $J_{1,2} = J_{1,5} = 5.1$ Hz, 1 H, 1-H), 6.17 (d, $J_{5,\rm NH} = 8.5$ Hz, 1 H, NH) ppm. HRMS: calcd. for C $_{17}$ H $_{25}$ NO $_{7}$ 355.1631; found 355.1612 [M⁺].
- **2,3-***O*-Cyclohexylidene Derivative of (1*S*,2*R*,3*S*,4*R*)-5-Acetamido-1,4-di-*O*-methyl-1,2,3,4-cyclopentanetetrol (33): Diol 31 (76 mg, 0.28 mmol) was methylated with silver oxide (194 mg, 0.837 mmol) and iodomethane (1.74 mL, 27.9 mmol) in acetonitrile (1.0 mL) at 80 °C for 5 h. The product was purified by chromatography on a

column of silica gel (3 g, acetone/toluene, 1:5) to give ether **33** (57 mg, 68%) as a syrup. TLC (acetone/toluene, 2:1): $R_{\rm f} = 0.62$. $[a]_{\rm D}^{\rm 122} = -31$ (c = 1.4, CHCl₃). 1 H NMR (300 MHz, CDCl₃): $\delta = 1.25-1.81$ (m, 10 H, C₆H₁₀), 3.44, 3.44 (2 s, each 3 H, 2×Me), 3.62 (s, 1 H, 4-H), 3.88 (t, $J_{1,2} = J_{1,5} = 4.9$ Hz, 1 H, 1-H), 4.41 (d, $J_{2,3} = 4.9$ Hz, 1 H, 3-H), 4.44 (dd, $J_{1,5} = 4.9$, $J_{5,\rm NH} = 7.1$ Hz, 1 H, 5-H), 4,73 (t, $J_{1,2} = J_{2,3} = 4.9$ Hz, 1 H, 2-H), 6.34 (d, $J_{5,\rm NH} = 7.1$ Hz, 1 H, NH) ppm. HRMS: calcd. for C₁₅H₂₆NO₅ 300.1811; found 300.1816 [M⁺].

(1*S*,2*R*,3*S*,4*R*)-5-Acetamido-2,3-di-*O*-acetyl-1,4-di-*O*-methyl-1,2,3,4-cyclopentanetetrol (15a): Compound 33 (54 mg, 0.18 mmol) was hydrolysed with 60% aqueous acetic acid at 80 °C for 2 h. The product was acetylated in the usual manner, and the acetyl derivative was purified by chromatography on a column of silica gel (3 g, acetone/toluene, 1:3) to give the di-*O*-acetyl derivative 15a (46.5 mg, 86%) as a syrup. TLC (acetone/toluene, 1:1): $R_{\rm f} = 0.39$. [a] $_{\rm f}^{22} = -44$ (c = 1.2, CHCl₃). $^{1}{\rm H}$ NMR (300 MHz, CDCl₃): $\delta = 2.06$, 2.06, 2.14 (3 s, each 3 H, 3×Ac), 3.37, 3.45 (2 s, each 3 H, 2×Me), 3.72 (dd, $J_{4,5} = 3.7$, $J_{3,4} = 5.6$ Hz, 1 H, 4-H), 3.89 (dd, $J_{1,2} = 3.6$, $J_{1,5} = 6.8$ Hz, 1 H, 1-H), 4.44 (ddd, $J_{4,5} = 3.7$, $J_{1,5} = 6.8$, $J_{5,\rm NH} = 8.5$ Hz, 1 H, 5-H), 5.02 (t, $J_{2,3} = J_{3,4} = 5.6$ Hz, 1 H, 3-H), 5.48 (dd, $J_{1,2} = 3.6$, $J_{2,3} = 5.6$ Hz, 1 H, 2-H), 6.05 (d, $J_{5,\rm NH} = 8.5$ Hz, 1 H, NH) ppm. HRMS: calcd. for $C_{13}H_{22}NO_7$ 304.1396; found 304.1402 [M⁺].

2,3-O-Cyclohexylidene Derivative of (1R,2S,3S,4S,5R)-5-Acetamido-1-O-triethylsilyl-1,2,3,4-cyclopentanetetrol (34): To a solution of D-17^[24] (114 mg, 0.254 mmol) in CH₂Cl₂ (1.5 mL) were added 2,6-lutidine (89 μ L, 0.763 mmol) and triethylsilyl triflate (86 µL, 0.38 mmol) at 0 °C under argon, and the mixture was stirred at 0 °C for 30 min. Saturated aqueous NaHCO₃ (20 mL) was added, and the mixture was extracted with chloroform (3×10) mL). The extracts were washed with water, dried, and the solvents were evaporated. The residual product was treated with 1 m methanolic sodium methoxide (50 µL) in CH₂Cl₂ (1.0 mL) at 0 °C for 30 min. After neutralization with aqueous acetic acid, an insoluble material was removed by filtration and washed with acetone. The filtrate and washings were combined and the solvents were evaporated to dryness. The residue was purified by chromatography on a column of silica gel (10 g, acetone/toluene, 1:4) to give the alcohol **34** (66 mg, 68%) as a syrup. TLC (acetone/toluene, 1:1): $R_f = 0.48$. $[a]_{D}^{21} = +25 \ (c = 1.0, \text{CHCl}_{3}).$ H NMR (300 MHz, CDCl₃): $\delta =$ 0.63 [dd, J = 8.1, $J_{gem} = 15.1$ Hz, 6 H, Si(C H_2 CH₃)₃], 0.97 [t, J =8.1 Hz, 9 H, $Si(CH_2CH_3)_3$], 1.36–1.83 (m, 10 H, C_6H_{10}), 2.04 (s, 3 H, Ac), 3.06 (d, $J_{4,OH}$ = 10.0 Hz, 1 H, OH), 4.00 (ddd, $J_{3,4}$ = $J_{4,5}$ = 6.6, $J_{4,OH}$ = 10.0 Hz, 1 H, 4-H), 4.08 (t, $J_{1,2}$ = $J_{1,5}$ = 4.4 Hz, 1 H, 1-H), 4.15 (ddd, $J_{1,5} = 4.4$, $J_{4,5} = 6.6$, $J_{5,NH} = 7.8$ Hz, 1 H, 5-H), 4.44 (dd, $J_{1,2}$ = 4.4, $J_{2,3}$ = 6.6 Hz, 1 H, 2-H), 4.53 (t, $J_{2,3}$ = $J_{3,4}$ = 6.6 Hz, 1 H, 3-H), 6.37 (d, $J_{5.NH}$ = 7.8 Hz, 1 H, NH) ppm. HRMS: calcd. for C₁₉H₃₅NO₅Si 385.2285; found 385.2284 [M⁺].

2,3-*O*-Cyclohexylidene Derivative of (1R,2S,3S,4S,5R)-5-Acetamido-1-*O*-methyl-4-*O*-triethylsilyl-1,2,3,4-cyclopentanetetrol (35): To a solution of 34 (57 mg, 0.15 mmol) in acetonitrile (1.0 mL) were added silver oxide (51 mg, 0.22 mmol) and a large excess of iodomethane (0.9 mL, 14.7 mmol), and the suspension was refluxed at 80 °C overnight. After cooling, an insoluble material was removed by filtration, and the filtrate was concentrated. The residue was purified by chromatography on a column of silica gel (3 g, acetone/toluene, 1:6) to give the methyl ether 35 (57 mg, 97%) as a syrup. TLC (acetone/toluene, 1:1): $R_f = 0.56$. $[a]_{2}^{D1} = +8.3$ (c = 1.0, CHCl₃). 1 H NMR (300 MHz, CDCl₃): $\delta = 0.63$ [dd, J = 8.1, $J_{gem} = 15.6$ Hz, 6 H, Si(CH₂CH₃)₃], 0.97 [t, J = 8.1 Hz, 9 H, Si(CH₂CH₃)₃], 1.32–1.84 (m, 10 H, C₆H₁₀), 2.00 (s, 3 H, Ac), 3.44

(t, $J_{3,4} = J_{4,5} = 5.4$ Hz, 1 H, 4-H), 3.46 (s, 3 H, Me), 3.92 (t, $J_{1,2} = J_{1,5} = 5.4$ Hz, 1 H, 1-H), 4.44 (t, $J_{1,2} = J_{2,3} = 5.4$ Hz, 1 H, 2-H), 4.60 (t, $J_{2,3} = J_{3,4} = 5.4$ Hz, 1 H, 3-H), 4.62 (ddd, $J_{1,5} = J_{4,5} = 5.4$, $J_{5,\mathrm{NH}} = 9.8$ Hz, 1 H, 5-H), 6.27 (d, $J_{5,\mathrm{NH}} = 9.8$ Hz, 1 H, NH) ppm. HRMS: calcd. for $\mathrm{C}_{20}\mathrm{H}_{37}\mathrm{NO}_5\mathrm{Si}$ 399.2441; found 399.2427 [M⁺].

(1*R*,2*R*,3*S*,4*S*,5*R*)-5-Acetamido-1,2,3-tri-*O*-acetyl-4-*O*-methyl-1,2,3,4-cyclopentanetetrol (16a): A mixture of 35 (10 mg, 25 μmol) and aqueous acetic acid (60%, 2.0 mL) was stirred at 80 °C for 9 h, and then coconcentrated with ethanol and toluene. The residue was treated with acetic anhydride (0.5 mL) and pyridine (1.0 mL) at room temperature overnight. The reaction mixture was concentrated, and the residue was purified by chromatography on a column of silica gel (0.7 g, acetone/toluene, 1:3) to give the acetyl derivative 16a (7.3 mg, 87%) as a syrup. TLC (acetone/toluene, 1:1): $R_{\rm f} = 0.41$. [a] $^{22}_{\rm D} = +36$ (c = 0.4, CHCl₃). 1 H NMR (300 MHz, CDCl₃): $\delta = 2.05$, 2.08, 2.09, 2.14 (4 s, each 3 H, 4×Ac), 3.40 (s, 3 H, Me), 3.86 (m, 1 H, 4-H), 4,74 (m, 1 H, 5-H), 5.22–5.29 (m, 3 H, 1-H, 2-H, 3-H), 5.97 (d, $J_{5,\rm NH} = 9.0$ Hz, 1 H, NH) ppm. HRMS: calcd. for C₁₄H₂₂NO₈ 332.1345; found 332.1331 [M⁺].

(15,25,3R,4R)-4-Amino-1,2,3-cyclopentanetriol (11): A mixture of 11a (8.3 mg, 28 µmol) and 2 M hydrochloric acid (1 mL) was stirred at 80 °C for 1 h, and the solvents were evaporated to dryness. The residual product was purified by chromatography on a column of Dowex-50 W × 2 (H⁺) resin (1 mL, 1 M aqueous ammonia) to give the free base 11 (3.7 mg, ca. 100%). TLC (H₂O/AcOH/tBuOH, 1:1:4): $R_{\rm f} = 0.33$. $[a]_{\rm D}^{24} = +46$ (c = 0.4, MeOH). ¹H NMR (300 MHz, D₂O): $\delta = 1.46$ (ddd, $J_{1,5a} = 6.1$, $J_{4,5a} = 6.8$, $J_{\rm gem} = 14.6$ Hz, 1 H, 5a-H), 2.35 (ddd, $J_{1,5b} = 7.6$, $J_{4,5a} = 6.8$, $J_{4,5b} = 8.5$ Hz, 1 H, 4-H), 3.80 (t, $J_{1,2} = J_{2,3} = 4.4$ Hz, 1 H, 2-H), 3.87 (dd, $J_{2,3} = 4.4$, $J_{3,4} = 5.6$ Hz, 1 H, 3-H), 3.97 (ddd, $J_{1,5b} = 7.6$ Hz, 1 H, 1-H) ppm. HRMS: calcd. for $C_5H_{12}NO_3$ 134.0817; found 134.0798 [M⁺].

(1*R*,2*R*,3*R*,4*S*,5*R*)-4-Amino-5-thio-1,2,3-cyclopentanetriol (12): Compound 12a (8.9 mg) was hydrolyzed as in the preparation of 11, and the product was purified on a column of Dowex-50 W × 2 (H⁺) resin (1 mL, 1 m aqueous ammonia) to give 12 (2.9 mg, 74%) as a syrup. TLC (H₂O/AcOH/*t*BuOH, 1:1:2): $R_f = 0.24$. [a] $_0^2 = +66$ (c = 0.1, MeOH). $_1^1$ H NMR (300 MHz, D₂O): $\delta = 3.12$ (dd, $J_{1,5} = 5.6$, $J_{4,5} = 7.8$ Hz, 1 H, 5-H), 3.88–3.98 (m, 3 H, 1-H, 2-H, 3-H), 3.95 (m, 1 H, 4-H) ppm. HRMS: calcd. for C₅H₁₃NO₃S 167.0616; found 167.0633 [M⁺].

(1*R*,2*R*,3*R*,4*S*,5*R*)-4-Amino-5-ethylthio-1,2,3-cyclopentanetriol (13): Compound 13a (5.9 mg, 16 μmol) was hydrolyzed as in the preparation of 11, and the product was purified by a column of Dowex-50 W × 2 (H⁺) resin (1 mL, 1 м aqueous ammonia) to give 13 (2.9 mg, ca. 100%) as a syrup. TLC (MeOH/chloroform, 1:2): R_f = 0.23. [a]_D²⁴ = -54 (c = 0.15, MeOH). ¹H NMR (300 MHz, D₂O): δ = 1.11 (t, J = 7.3 Hz, 3 H, CH₂CH₃), 2.54 (br. q, J = 7.3 Hz, 2 H, CH₂CH₃), 2.84 (dd, $J_{3,4}$ = 6.3, $J_{4,5}$ = 7.8 Hz, 1 H, 4-H), 3.00 (dd, $J_{1,5}$ = 5.6, $J_{4,5}$ = 7.8 Hz, 1 H, 5-H), 3.82 (dd, $J_{2,3}$ = 5.4, $J_{3,4}$ = 6.3 Hz, 1 H, 3-H), 3.90 (t, $J_{1,2}$ = $J_{2,3}$ = 5.4 Hz, 1 H, 2-H), 3.96 (dd, $J_{1,2}$ = 5.4, $J_{1,5}$ = 5.6 Hz, 1 H, 1-H) ppm. HRMS: calcd. for C₇H₁₅NO₃S 193.0773; found 193.0781 [M⁺].

(1*R*,2*R*,3*R*,4*R*,5*S*)-5-Amino-4-*O*-methyl-1,2,3,4-cyclopentanetetrol (14): Compound 14a (9.2 mg, 28 µmol) was treated with 2 M aqueous barium hydroxide (1 mL) at 90 °C for 5 h. After neutralization with carbon dioxide, the reaction mixture was filtered through a pad of Celite and the filtrate was concentrated to dryness. The product was purified on a column of Dowex-50 W×2 (H⁺) resin (1 mL, 1% aqueous ammonia) to give the free base 14 (1.9 mg, 42%) as a syrup. TLC (H₂O/AcOH/BuOH, 1:1:2): $R_f = 0.46$. [a]²⁰ = +53

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(c = 0.1, MeOH). ¹H NMR (300 MHz, D₂O): $\delta = 2.92$ (m, 1 H, 5-H), 3.32 (s, 3 H, Me), 3.48 (m, 1 H, 4-H), 3.81 (m, 1 H, 3-H), 3.89 (m, 2 H, 1-H, 2-H) ppm. HRMS: calcd. for C₆H₁₄NO₄ 164.0923; found 164.0922 [M⁺].

(1R,2S,3R,4R)-5-Amino-1,4-di-O-methyl-1,2,3,4-cyclopentanetetrol (15): Compound 15a (7.4 mg, 24 μmol) was treated with 1 M aqueous barium hydroxide (1 mL) at 90 °C overnight. After neutralization with CO₂, an insoluble material was removed by filtration through a Celite pad, and the filtrate was concentrated to dryness. The residue was purified by chromatography on a column of Dowex-50 W \times 2 (H⁺) resin (1 mL, 1 M aqueous ammonia) to give the di-O-methyl ether 15 (4.3 mg, ca. 100%) as a syrup. TLC (H₂O/ AcOH/BuOH, 1:1:2): $R_f = 0.41$. $[a]_D^{22} = +23$ (c = 0.3, MeOH). ¹H NMR (300 MHz, D₂O): δ = 3.13 (dd, $J_{4,5}$ = 5.4, $J_{1,5}$ = 6.3 Hz, 1 H, 5-H), 3.31, 3.32 (2 s, each 3 H, $2 \times Me$), 3.50 (t, $J_{3,4} = J_{4,5} =$ 5.4 Hz, 1 H, 4-H), 3.58 (dd, $J_{1,2} = 3.7$, $J_{1,5} = 6.3$ Hz, 1 H, 1-H), 3.77 (t, $J_{2,3} = J_{3,4} = 5.4$ Hz, 1 H, 3-H), 4.03 (dd, $J_{1,2} = 3.7$, $J_{2,3} =$ 5.4 Hz, 1 H, 2-H) ppm. HRMS: calcd. for C₇H₁₆NO₄ 178.1079; found 178.1087 [M+].

(1S,2R,3R,4R,5S)-5-Amino-1-O-methyl-1,2,3,4-cyclopentanetetrol (16): Compound 16a (57 mg, 17 μmol) was treated with 1 м aqueous barium hydroxide (1 mL) at 90 °C overnight. After neutralization with carbon dioxide, the mixture was filtered through a pad of Celite which was washed with water thoroughly. The filtrate and washings were concentrated and the residue was purified by chromatography on a column of Dowex-50 W×2 (H⁺) resin (1 mL, 1 M aqueous ammonia) to give the amine 16 (2.8 mg, ca. 100%) as a syrup. TLC (H₂O/AcOH/BuOH, 1:1:3): $R_f = 0.19$. $[a]_D^{20} = -16$ (c = 0.14, MeOH). ¹H NMR (300 MHz, D_2O): δ = 3.32 (m, 4 H, 5-H, Me), 3.60 (dd, $J_{1,2}$ = 4.9, $J_{1,5}$ = \approx 5 Hz, 1 H, 1-H), 3.81 (t, $J_{2,3}$ = $J_{3,4}$ = 5.1 Hz, 1 H, 3-H), 3.92 (t, $J_{3,4}$ = $J_{4,5}$ = 5.1 Hz, 1 H, 4-H), 3.99 (dd, $J_{1,2}$ = 4.9, $J_{2,3}$ = 5.1 Hz, 1 H, 2-H) ppm. HRMS: calcd. for C₁₄H₂₂NO₈ 164.0923; found 164.0915 [M⁺].

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- [4] S. Ogawa, Y. Yuming, J. Chem. Soc., Chem. Commun. 1991,
- [5] S. B. King, B. Ganem, J. Am. Chem. Soc. 1991, 113, 5089-
- S. B. King, B. Ganem, J. Am. Chem. Soc. 1994, 116, 562–571.
- S. Knapp, T. G. Murali Dhar, J. Org. Chem. 1991, 56, 4096-
- B. M. Trost, D. L. Van Vranken, J. Am. Chem. Soc. 1991, 113, [8] 5089-5090.
- B. Ganem, Carbohydrate Mimics (Ed.: Y. Chapleur), Wiley-VCH, Weinheim, **1998**, pp. 239–258.
- [10] For a comprehensive review of synthetic and biological features of the mannostatins, see: A. Berecibar, C. Grandjean, A. Siriwardena, Chem. Rev. 1999, 99, 779-844.
- [11] In this paper the nomenclature of aminocyclopentitols and derivatives follows the IUPAC-IUB 1973 Recommendations for Cyclitols (Pure Appl. Chem. 1974, 37, 285-297). In the Exp. Sect., the (R,S) notation is generally used instead to describe absolute configurations.
- [12] G. Legler, Adv. Carbohydr. Chem. Biochem. 1990, 48, 319-384.
- [13] G. Legler, Carbohydrate Mimics (Ed.: Y. Chapleur), Wiley-VCH, Weinheim, 1998, pp. 463-490.
- [14] G. C. Lock, C. H. Fotsch, C. H. Wong, Acc. Chem. Res. 1993, *26*, 182–190.
- [15] D. A. Winkler, G. Holan, J. Med. Chem 1989, 32, 2084-2089.
- [16] D. A. Winkler, J. Med. Chem. 1996, 39, 4332-4334.
- [17] S. Ogawa, T. Morikawa, Bioorg. Med. Chem. Lett. 1999, 9, 1499-1504.
- [18] S. Ogawa, T. Morikawa, Eur. J. Org. Chem. 2000, 1759–1765.
- [19] C. Uchida, H. Kimura, S. Ogawa, Bioorg. Med. Chem. 1997, 5, 921–939.
- [20] S. Ogawa, K. Washida, Eur. J. Org. Chem. 1998, 1929–1934.
- [21] S. J. Cho, R. Ling, A. Kim, P. S. Mariano, J. Org. Chem. 2000, 65, 1574-1577.
- [22] S. B. King, B. Ganem, J. Am. Chem. Soc. 1994, 116, 562–564.
- [23] Y. Nishimura, Y. Umezawa, H. Adachi, S. Kondo, T. Takeuchi, J. Org. Chem. 1996, 61, 480-488.
- [24] T. Suami, K. Tadano, S. Nishiyama, F. W. Lichtenthaler, J. Org. Chem. 1973, 38, 3691-3696.
- [25] C. Uchida, T. Yamagishi, S. Ogawa, J. Chem. Soc., Perkin Trans. 1 1994, 589-602.
- [26] S. Ogawa, H. Kimura, C. Uchida, T. Ohashi, J. Chem. Soc., Perkin Trans. 1 1995, 1695–1705.
- [27] H. Suzuki, S.-C. Li, Y.-T. Li, J. Biol. Chem. 1970, 245, 781-
- [28] L. G. Dickson, E. Leroy, J.-L. Reymond, Org. Biomol. Chem. **2004**, 2, 1217–1226.
- [29] R. A. Farr, N. Peet, M. S. Kang, Tetrahedron Lett. 1990, 31, 7109-7112.
- [30] M. Kleban, P. Hilgers, J. N. Greul, R. D. Kugler, J. Li, S. Picasso, P. Vogel, V. Jäger, ChemBioChem. 2001, 365–368.
- [31] R. C. Bernotas, G. Papandreou, J. Urbach, B. Ganem, Tetrahedron Lett. 1990, 31, 3393-3396.

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^[1] S. Ogawa, T. Morikawa, Bioorg. Med. Chem. Lett. 2000, 10,

T. Aoyagi, T. Yamamoto, K. Kojiri, H. Morishima, M. Nagai, M. Hamada, T. Takeuchi, H. Umezawa, J. Antibiot. 1989, 42, 883–889.

^[3] H. Morishima, K. Kojiri, T. Yamamoto, T. Aoyagi, H. Nakamura, Y. Iitaka, J. Antibiot. 1989, 42, 1008–1011.